

***Aesculus* species: a review on biologically active compounds and their possible applications**

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Abstract

Horse chestnut (*Aesculus hippocastanum* L.) is a specific tree of Eastern Europe, including Balkan Mountains. It is an ornamental tree, but its seeds are valuable for their important content of triterpenoid saponins, especially β -aescin (or β -escin). Other biologically active compounds found in *A. hippocastanum* are flavonoids, anthocyanidins and their glycosides. In this review, the biologically active compounds from *Aesculus* species, especially *A. hippocastanum*, have been reported. The classification, extraction and purification methods of biologically active compounds from these species were emphasized. Moreover, the main biological activities of both *Aesculus* extracts and purified compounds, including the treatment of chronic venous insufficiency, vascular protection and venotonic effect, as well as anti-inflammatory and antioxidant activities, were reviewed.

Keywords: *Aesculus hippocastanum* L., horse chestnut, triterpenoids, saponins, triterpene glycosides, aescins, escins, β -aescin, β -escin, chronic venous insufficiency

1. Introduction

Aesculus hippocastanum L. (horse chestnut or European horsechestnut) is one of the *Aesculus* species originated from Europe, but it is cultivated all over the world. The tree belongs to the genus *Aesculus* and is valuable even as ornamental tree or for its useful seeds [1-4]. It was used for the treatment of chronic venous inefficiency, varicose veins, inflammation of the veins, haemorrhoids, diarrhoea, fever, prostate disorders, rheumatism and neuralgia [5-8]. Among seeds, other parts of the horse chestnut tree is used for above mentioned applications, such as bark and leaves, all as extracts or other preparations [8, 9]. There are many formulations based on *A. hippocastanum* such as external elastic compresses, sprain, bruise, topical gels, as well as shampoos, foam baths, creams or lotions for skin care and toiletries [9].

There are other *Aesculus* species that were studied both for their composition regarding biologically active compounds and various biological activities. It is the case of *A. californica* (Spach) Nutt. (California buckeye or California horse-chestnut, United States of America), *A. chinensis* Bunge (China), *A. glabra* Willd. (Ohio buckeye or American buckeye, Midwest of the USA), *A. indica* Colebr. (Indian horse chestnut or *Han dun*, temperate regions of Asia), *A. pavia* L. (red buckeyes or scarlet buckeye, originated from the southeast of the USA), *A. sylvatica* W. Bartram (eastern USA), and *A. turbinata* Blume (Japanese horse chestnut or *Tochinomi*, Japan.) [10-16]. An up-to-date survey on the biologically active compound composition and the possible applications is presented below.

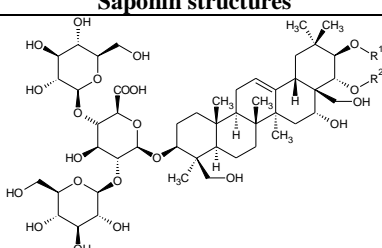
2. Biologically active compounds from *Aesculus* species

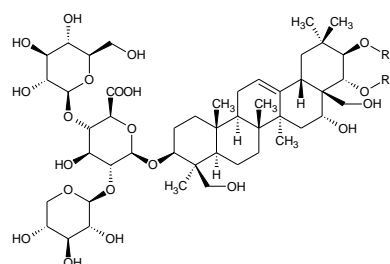
The *A. hippocastanum* seeds are the most important part of the tree. Carbohydrates are the most concentrated components of seeds. They comprise of water-soluble and non-water-soluble saccharides (6.6% and 18.1%, respectively), lignin 9.7%, hemicellulose and cellulose (10.8% and 3.6%, respectively). Proteins and lipids were found in concentrations of approximately 34.4% and 13.1%, while ash was at 3% [17]. Moreover, Mediterranean varieties having white or pink flowers had a protein content of 2.64% and 1.82%, while the lipid contents were 4.13% and 5.10%, respectively [18].

By far, the most important biologically active compounds are saponins with aescins (or escins) as representatives. They provide the main biological activities of the *Aesculus* species. Flavonoids and anthocyanins (and proanthocyanins) are antioxidant compounds that have also been found in various parts of the tree. Fatty acid glycerides are mainly separated from *Aesculus* seeds, while carbohydrates are the major constituents [3-5, 8, 9, 17, 19, 20].

Saponins or triterpenoid glycosides have steroid-based aglycones. Aglycones have saccharide moieties attached through one or more hydroxyl groups. *Aesculus* species have various contents of such saponins, the most important being aescin Ia (with the synonyms β -aescin or β -escin), as well as Ib, Iia, and Iib. They were found in *A. hippocastanum* and *A. turbinata* [21-26]. Desacylaescins have also found in these species. On the other hand, aesculosides, vaccarosides, and paviosides have also been found in *A. galabra*, *A. sylvatica*, *A. californica*, *A. pavia* and *A. turbinata* [10, 12, 14, 15, 27-29]. A survey on the saponin structures found in *Aesculus* species is presented in Table 1.

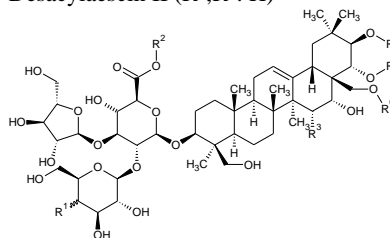
Table 1. Structures of the main saponins found in *Aesculus* species

Saponin structures	<i>Aesculus</i> species
	<i>A. hippocastanum</i> , <i>A. turbinata</i> [21-26]
Aescin Ia (R ¹ : tigloyl, R ² : acetyl)	
Aescin Ib (R ¹ : angeloyl, R ² : acetyl)	
Desacylaescin I (R ¹ , R ² : H)	



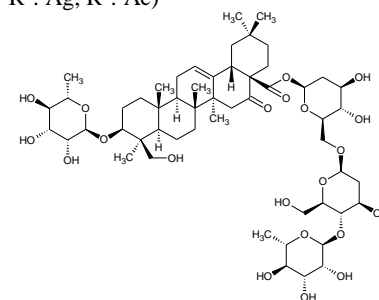
Aescin Iia (R¹: Tg, R²: Ac)
Aescin Iib (R¹: Ag, R²: Ac)
Desacylaescin II (R¹, R²: H)

A. hippocastanum,
A. turbinata
[21-26]



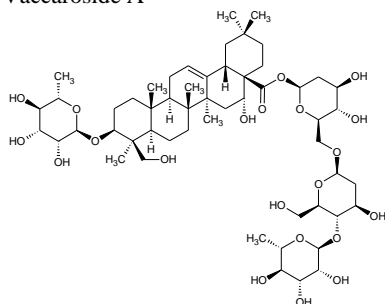
Aesculoside G1 (R¹: β OH, R^{2,5}: H, R³: OH, R⁴: Tg, R⁶: Ac)
Aesculoside G2 (R¹: β OH, R^{2,5}: H, R³: OH, R⁴: Ag, R⁶: Ac)
Aesculoside G3 (R¹: β OH, R^{2,3,5}: H, R⁴: Tg, R⁶: Ac)
Aesculoside G4 (R¹: β OH, R^{2,3,5}: H, R⁴: Ag, R⁶: Ac)
Aesculoside G5 (R¹: α OH, R^{2,5,6}: H, R³: OH, R⁴: Ag)
Aesculoside G6 (R¹: β OH, R^{2,6}: H, R³: OH, R⁴: Tg, R⁵: Ag)
Aesculoside G7 (R¹: β OH, R^{2,6}: H, R³: OH, R⁴: Ag, R⁵: Tg)
Aesculoside G8 (R¹: β OH, R^{2,3,6}: H, R⁴: Tg, R⁵: Tg)
Aesculoside G9 (R¹: β OH, R^{2,3,6}: H, R⁴: Tg, R⁵: Ag)
Aesculoside G10 (R¹: β OH, R^{2,3,6}: H, R⁴: Ag, R⁵: Tg)
Aesculoside G11 (R¹: β OH, R²: Me, R^{3,6}: H, R⁴: Ag, R⁵: Tg)
Aesculoside G12 (R¹: β OH, R²: Me, R^{3,6}: H, R⁴: Ag, R⁵: Ag)
Aesculoside G13 (R¹: α OH, R^{2,6}: H, R³: OH, R⁴: Tg, R⁵: Ac)
Aesculoside G14 (R¹: α OH, R^{2,6}: H, R³: OH, R⁴: Ag, R⁵: Ac)
Aesculoside G15 (R¹: α OH, R^{2,3,6}: H, R⁴: Tg, R⁵: Ac)
Aesculoside G16 (R¹: α OH, R^{2,3,6}: H, R⁴: Ag, R⁵: Ac)

A. galabra,
A. sylvatica,
A. californica,
A. pavia
[10, 12, 14, 15, 27]



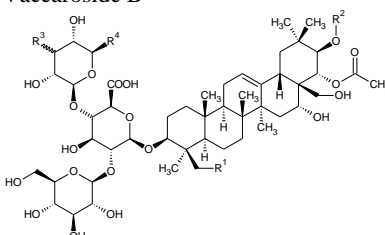
A. pavia
[28]

Vaccaroside A



A. pavia
[28]

Vaccaroside B



A. pavia
[29]

Pavioside A (R¹: H, R²: Tg, R³: αOH, R⁴: CH₂OH)

Pavioside B (R¹: H, R²: Ag, R³: αOH, R⁴: CH₂OH)

Pavioside C (R¹: H, R²: Tg, R³: βOH, R⁴: CH₂OH)

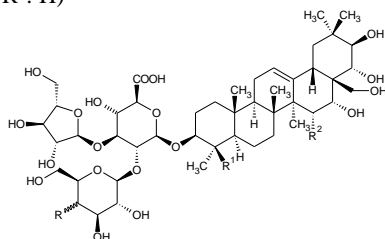
Pavioside D (R¹: H, R²: Ag, R³: βOH, R⁴: CH₂OH)

Pavioside E (R¹: H, R²: Tg, R³: αOH, R⁴: H)

Pavioside F (R¹: H, R²: Ag, R³: αOH, R⁴: H)

Pavioside G (R¹: OH, R²: Tg, R³: αOH, R⁴: H)

Pavioside H (R¹: OH, R²: Ag, R³: αOH, R⁴: H)



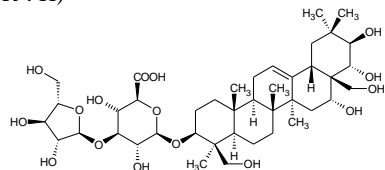
A. turbinata
[27]

Aesculoside Ia (R: βOH, R¹: Me, R²: OH)

Aesculoside Ib (R: αOH, R¹: CH₂OH, R²: OH)

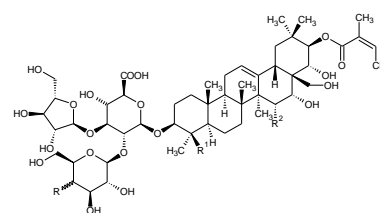
Aesculoside Ic (R: βOH, R¹: Me, R²: H)

Aesculoside Id (R: αOH, R¹: CH₂OH, R²: H)



Aesculoside Ie

A. turbinata
[27]

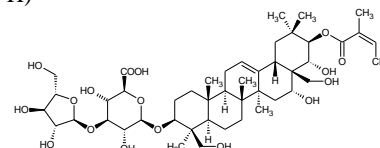


A. turbinata
[27]

Aesculoside IIa (R: βOH, R¹: Me, R²: OH)

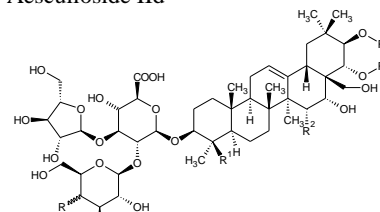
Aesculoside IIb (R: αOH, R¹: CH₂OH, R²: H)

Aesculoside IIc (R: βOH, R¹: Me, R²: H)



A. turbinata
[27]

Aesculoside IIc

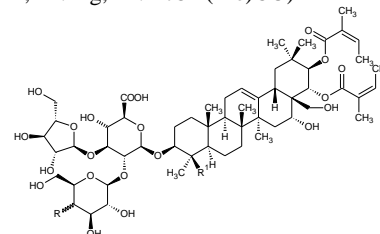


A. turbinata
[27]

Aesculoside IVa (R: βOH, R¹: Me, R²: OH, R³: Ag, R⁴: EtCH(Me)CO)

Aesculoside IVb (R: αOH, R¹: CH₂OH, R²: H, R³: Ag, R⁴: EtCH(Me)CO)

Aesculoside IVc (R: βOH, R¹: Me, R²: H, R³: Ag, R⁴: EtCH(Me)CO)



A. turbinata
[27]

Aesculoside derivative a (R: αOH, R¹: CH₂OH)

Aesculoside derivative b (R: βOH, R¹: Me)

* Tg=tigloyl: *trans*-MeCH=C(Me)CO; Ag=angeloyl: *cis*-MeCH=C(Me)CO; Ac=acetyl: MeCO

Flavonoids are valuable compounds that have antioxidant activity. Some compounds from this class was found in *Aesculus* species, as well as catechins and procyanidins (especially trimers). Examples of these structures are presented in Tables 2 and 3.

Table 2. Structures of the main flavonoids found in *Aesculus* species

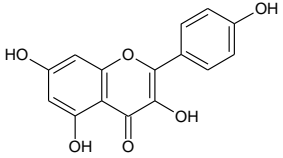
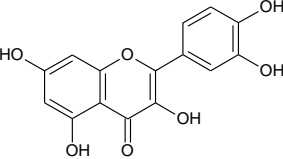
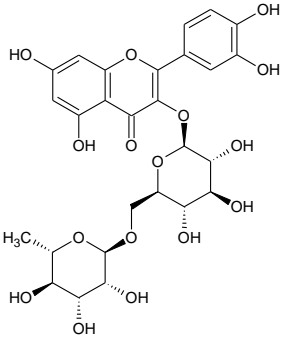
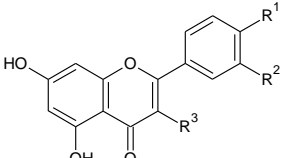
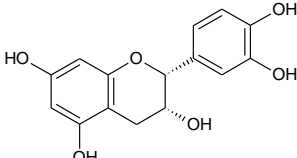
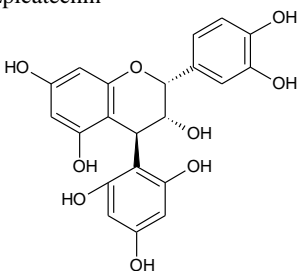
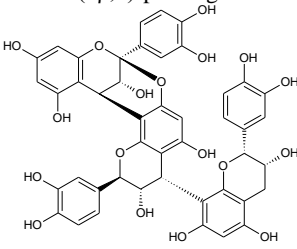
Flavonoid structures	<i>Aesculus</i> species
	<i>A. hippocastanum</i> [30]
Kaemferol	
	<i>A. hippocastanum</i> [30]
Quercetin	
	<i>A. hippocastanum</i> [30]
Rutin	
	<i>A. hippocastanum</i> , <i>A. chinensis</i> , <i>A. turbinata</i> [16, 19, 31]
Flavonoid derivative 1 (R ¹ : OH, R ² : Glc, R ³ : Xyl-Rha-Glc)	
Flavonoid derivative 2 (R ¹ : OH, R ² : Glc, R ³ : Xyl(1-2)Glc)	
Flavonoid derivative 3 (R ^{1,2} : OH, R ³ : Xyl(1-2)[Glc(1-3)]Glc)	
Flavonoid derivative 4 (R ^{1,2} : OH, R ³ : Xyl(1-2)Glc)	
Flavonoid derivative 5 (R ¹ : OH, R ² : Glc-nicotinoyl, R ³ : Xyl(1-2)Glc)	
Flavonoid derivative 6 (R ¹ : OH, R ² : Glc-3-hydroxy-2-oxoindolin-3-ilacetil, R ³ : Xyl(1-2)Glc)	
Flavonoid derivative 7 (R ¹ : H, R ² : OH, R ³ : Xyl(1-2)[Glc(1-3)]Glc)	
Flavonoid derivative 8 (R ¹ : H, R ² : OH, R ³ : Xyl(1-2)Glc)	
Flavonoid derivative 9 (R ¹ : OMe, R ² : OH, R ³ : Xyl(1-2)[Glc(1-3)]Glc)	
Flavonoid derivative 10 (R ¹ : OMe, R ² : Glc, R ³ : Xyl(1-2)[Glc(1-3)]Glc)	
Flavonoid derivative 11 (R ^{1,2} : OH, R ³ : Glc-Rha)	
Flavonoid derivative 12 (R ¹ : OH, R ² : 2-oxoindol-3-ylacetyl, R ³ : Xyl(1-2)Glc)	
Flavonoid derivative 13 (R ¹ : H, R ² : OH, R ³ : Glc-Rha)	

Table 3. Structures of some catechins and procyanidins found in *Aesculus* species

Catechin or procyanidin structures	<i>Aesculus</i> species
	<i>A. hippocastanum</i> [32]
(-)-Epicatechin	
	<i>A. hippocastanum</i> [33]
Epicatechin-(4β,2)-phloroglucinol	
	<i>A. hippocastanum</i> , <i>A. turbinata</i> [33, 34]
Epicatechin-(4β,8;2β,7)-catechin-(4β,8)-epicatechin	

Fatty acids, as triglycerides, were found in the seed oils of *A. hippocastanum*. The main compounds were oleic and linoleic acids (43.2-59.4% and <35.2%, respectively), but other monounsaturated fatty acids were identified at lower concentrations (gondoic, erucic and nervonic acids) [18, 30, 35].

The most concentrated compounds in *Aesculus* species are carbohydrates (amylose and amylopectin). However, they are less important from the biological activity point of view. There are some studies regarding the chemical changes and characteristics of *Aesculus* starch subjected to various processing conditions [13, 36-38].

Another interesting compound found in *A. hippocastanum* is plastoquinone-8, a structure resembling with coenzyme Q10 [39].

3. Biological activities of *Aesculus* species

The main biological activities of *Aesculus* species are due to the presence of saponins, especially aescins. They provide vascular and venotonic effects, as well as anti-inflammatory properties [5, 20]. On the other hand, the presence of flavonoids and procyanidins furnish antioxidant properties and

related biological activities. A survey on the main biological activities and applications of *Aesculus* compounds and extracts is presented in Table 4.

Table 4. The main biological activities and applications of bioactive compound and extracts from *Aesculus* species

<i>Aesculus</i> species	Biological activity
<i>A. hippocastanum</i> (standardized extract to 20% aescins)	Anti-inflammatory, decreasing platelet aggregation, increasing venous contractions and protecting venous endothelium relaxation, treating varicose veins, venous ulcers, leg-tiredness, swelling, and the hardening of the skin caused by lipodermatosclerosis [9]
<i>A. hippocastanum</i> (seed extract)	Treatment of chronic venous insufficiency [6]
<i>A. hippocastanum</i> (fresh seed extract)	Treatment of chronic venous insufficiency [7]
<i>A. hippocastanum</i> (various medicinal preparations)	Treatment of chronic venous insufficiency [8]
<i>A. hippocastanum</i> (flower extract)	Wound healing [40]
<i>A. hippocastanum</i> (seed extract and β-aescin)	Virucidal, antiviral and immunomodulatory activities against viruses HSV-1, VSV and Dengue virus [41]
<i>A. hippocastanum</i> (aescins and desacetylaescins)	Anti-inflammatory effects [21]
<i>A. hippocastanum</i> (seeds)	Treatment of diabetic nephropathy [42]
<i>A. hippocastanum</i> (aescins)	Protective effects on endotoxin-induced liver injury [22]
<i>A. hippocastanum</i> (extracts)	Cosmetic skin-care products [4]
<i>A. hippocastanum</i> (seed extract)	Genotoxic and antioxidant activity [43]
<i>A. hippocastanum</i> (seed extract)	Increase the antioxidative defense system of the body and prevent lipid peroxidation [44]
<i>A. hippocastanum</i> (leaves extract)	Biosynthesis of antibacterial silver nanoparticles [45]
<i>A. hippocastanum</i> (bark extract)	Emulsion stabilizer [46]
<i>A. californica</i> (aesculosides)	Cytotoxicity to human non-small cell lung tumor (A549) [10]
<i>A. glabra</i> (aesculosides)	Cytotoxicity against A549 and PC-3 cancer cell lines [12]
<i>A. pavia</i> (saponosides)	Cytotoxicity against various carcinoma cell lines [14, 28, 29]
<i>A. turbinata</i> (aescin derivatives)	inhibitory activities on porcine epidemic diarrhea virus replication [47]
<i>A. turbinata</i> (seeds)	Antioxidant activities [16]
<i>A. turbinata</i> (aescins)	Inhibitory effect on pancreatic lipase [26]
<i>A. turbinata</i> (seed shells)	Anti-obesity effects [34]

4. Conclusions

In this review an up-to-date survey on the chemical composition, biological activities and applications of compounds and extracts from various parts of *Aesculus* species was performed. Saponins (e.g. aescins), flavonoids, catechins and procyanidins from *A. hippocastanum*, *A. galabra*, *A. sylvatica*, *A. californica*, *A. chinensis*, *A. pavia* and *A. turbinata* were emphasized. Moreover, the biological activities and applications of both purified compounds and *Aesculus* extracts were systematically presented. This review can be useful for identifying new possible applications of *Aesculus*-based formulations in the pharmaceutical, cosmetic and even food fields.

References

- Gilman, E.F.; Watson, D.G. *Aesculus hippocastanum* Horsechestnut. Institute of Food and Agricultural Sciences, University of Florida, *Fact Sheet*. **1993**, ST-61, 1-4.
- Kitagawa, J.; Yasuda, Y. Development and distribution of *Castanea* and *Aesculus* culture during the Jomon Period in Japan. *Quaternary International*. **2008**, *184*, 41-55. <http://dx.doi.org/10.1016/j.quaint.2007.09.014>.
- Ravazzi, C.; Caudullo, G. *Aesculus hippocastanum* in Europe: distribution, habitat, usage and threats. In: San-Miguel-Ayanz, J.; de-Rigo, D.; Caudullo, G.; Houston-Durrant, T.; Mauri, A., editors. European Atlas of Forest Tree Species. Luxembourg: Publ. Off. EU, **2016**. p. e017fc3+.
- Wilkinson, J.A.; Brown, A.M.G. Horse chestnut – *Aesculus hippocastanum*: potential applications in cosmetic skin-care products. *International Journal of Cosmetic Science*. **1999**, *21*, 437-447.
- Karatoprak, G.Ş. Horse chestnut. In: Nabavi, S.M.; Silva, A.S., editors. Nonvitamin and nonmineral nutritional supplements. London: Elsevier Inc., **2019**. p. 295-299. <https://doi.org/10.1016/B978-0-12-812491-8.00042-4>.
- Pittler, M.; Ernst, E. Horse chestnut seed extract for chronic venous insufficiency. *Archives of Dermatology*. **1998**, *134*, 1356-1360.
- Suter, A.; Bommer, S.; Rechner, J. Treatment of patients with venous insufficiency with fresh plant horse chestnut seed extract: A review of 5 clinical studies. *Advances in Therapy*. **2006**, *23*(1), 179-190.
- Zampieron, E.R. Horse chestnut (*Aesculus hippocastanum*) for venous insufficiency. *International Journal of Complementary & Alternative Medicine*. **2017**, *5*(3), 153. <http://dx.doi.org/10.15406/ijcam.2017.05.00153>.

9. McCune, L.M. A Review of the Antioxidant Actions of Three Herbal Medicines (*Crataegus monogyna*, *Ginkgo biloba*, and *Aesculus hippocastanum*) on the Treatment of Cardiovascular Diseases. In: Watson, R.R.; Preedy, V.R., editors. Bioactive food as dietary interventions for cardiovascular disease. Boston: Elsevier Inc.; **2013**. p. 243-253.
10. Yuan, W.; Wang, P.; Su, Z.; Wang, V.S.; Li, S. Cytotoxic triterpenoid saponins from husks of *Aesculus californica* (Spach) Nutt. *Phytochemistry*. **2013**, *90*, 95-105. <http://dx.doi.org/10.1016/j.phytochem.2013.02.003>.
11. Chen, J.; Li, W.; Yang, B.; Guo, X.; Lee, F.S.-C.; Wang, X. Determination of four major saponins in the seeds of *Aesculus chinensis* Bunge using accelerated solvent extraction followed by high-performance liquid chromatography and electrospray-time of flight mass spectrometry. *Analytica Chimica Acta*. **2007**, *596*, 273-280. <http://dx.doi.org/10.1016/j.aca.2007.06.011>.
12. Yuan, W.; Wang, P.; Deng, G.; Li, S. Cytotoxic triterpenoid saponins from *Aesculus glabra* Willd. *Phytochemistry*. **2012**, *75*, 67-77. <http://dx.doi.org/10.1016/j.phytochem.2011.11.012>.
13. Wani, I.A.; Jabeen, M.; Geelani, H.; Masoodi, F.A.; Saba, I.; Muzaffar, S. Effect of gamma irradiation on physicochemical properties of Indian Horse Chestnut (*Aesculus indica* Colebr.) starch. *Food Hydrocolloids*. **2014**, *35*, 253-263. <http://dx.doi.org/10.1016/j.foodhyd.2013.06.002>.
14. Zhang, Z.; Li, S. Cytotoxic triterpenoid saponins from the fruits of *Aesculus pavia* L. *Phytochemistry*. **2007**, *68*, 2075-2086. <http://dx.doi.org/10.1016/j.phytochem.2007.05.020>.
15. Yuan, W.; Wang, P.; Su, Z.; Gao, R.; Li, S. Triterpenoid saponins from *Aesculus sylvatica* W. Bartram. *Phytochemistry Letters*. **2015**, *14*, 111-114. <http://dx.doi.org/10.1016/j.phytol.2015.09.011>.
16. Kimura, H.; Ogawa, S.; Ishihara, T.; Maruoka, M.; Tokuyama-Nakai, S.; Jisaka, M.; Yokota, K. Antioxidant activities and structural characterization of flavonol O-glycosides from seeds of Japanese horse chestnut (*Aesculus turbinata* BLUME). *Food Chemistry*. **2017**, *228*, 348-355. <http://dx.doi.org/10.1016/j.foodchem.2017.01.084>.
17. Marton, G.; Bakáin, A. Research and development on an integrated and wastefree utilization of horse chestnut (*Aesculus hippocastanum*) seeds. *Bioresource Technology*. **1995**, *54*, 197-200.
18. Baraldi, C.; Bodecchi, L.M.; Cocchi, M.; Durante, C.; Ferrari, G.; Foca, G.; Grandi, M.; Marchetti, A.; Tassi, L.; Ulrici, A. Chemical composition and characterisation of seeds from two varieties (pure and hybrid) of *Aesculus hippocastanum*. *Food Chemistry*. **2007**, *104*, 229-236. <http://dx.doi.org/10.1016/j.foodchem.2006.11.032>.
19. Kapusta, I.; Janda, B.; Szajwaj, B.; Stochmal, A.; Piacente, S.; Pizza, C.; Franceschi, F.; Franz, C.; Oleszek, W. Flavonoids in horse chestnut (*Aesculus hippocastanum*) seeds and powdered waste water byproducts. *Journal of Agricultural and Food Chemistry*. **2007**, *55*, 8485-8490. <http://dx.doi.org/10.1021/jf071709t>.
20. Tiffany, N.; Ulbricht, C.; Bent, S.; Smith, M.; Dennehy, C.; Boon, H.; Basch, E.; Barrette, E.P.; Sollars, D.; Szapary, P. Horse Chestnut: A multidisciplinary clinical review. *Journal of Herbal Pharmacotherapy*. **2002**, *2*(1), 71-85.
21. Matsuda, H.; Li, Y.; Murakami, T.; Ninomiya, K.; Araki, N.; Yoshikawa, M.; Yamahara, J. Antiinflammatory effects of escins Ia, Ib, IIa, and IIb from horse chestnut, the seeds of *Aesculus hippocastanum* L. *Bioorganic & Medicinal Chemistry Letters*. **1997**, *7*(13), 1611-1616.
22. Jiang, N.; Xin, W.; Wang, T.; Zhang, L.; Fan, H.; Du, Y.; Li, C.; Fu, F. Protective effect of aescin from the seeds of *Aesculus hippocastanum* on liver injury induced by endotoxin in mice. *Phytomedicine*. **2011**, *18*, 1276-1284. <http://dx.doi.org/10.1016/j.phymed.2011.06.011>.
23. Apers, S.; Naessens, T.; Pieters, L.; Vlietinck, A. Densitometric thin-layer chromatographic determination of aescin in a herbal medicinal product containing *Aesculus* and *Vitis* dry extracts. *Journal of Chromatography A*. **2006**, *1112*, 165-170. <http://dx.doi.org/10.1016/j.chroma.2005.10.069>.
24. Profumo, P.; Caviglia, A.M.; Gastaldo, P. Formation of aescin glucosides by callus tissue from cotyledonary explants of *Aesculus hippocastanum* L. *Plant Science*. **1992**, *85*, 161-164.
25. Vanhaelen, M.; Vanhaelen-Fastre, R. Quantitative determination of biologically active constituents in medicinal plant crude extracts by thin-layer chromatography-densitometry. *Journal of Chromatography*. **1983**, *281*, 263-271.
26. Kimura, H.; Ogawa, S.; Jisaka, M.; Kimura, Y.; Katsube, T.; Yokota, K. Identification of novel saponins from edible seeds of Japanese horse chestnut (*Aesculus turbinata* Blume) after treatment with wooden ashes and their nutraceutical activity. *Journal of Pharmaceutical and Biomedical Analysis*. **2006**, *41*, 1657-1665. <http://dx.doi.org/10.1016/j.jpba.2006.02.031>.
27. Zhang, Z.; Li, S.; Zhang, S.; Gorenstein, D. Triterpenoid saponins from the fruits of *Aesculus pavia*. *Phytochemistry*. **2006**, *67*, 784-794. <http://dx.doi.org/10.1016/j.phytochem.2006.01.017>.
28. Sun, Z.-L.; Zhang, M.; Wu, Y.; Wan, A.-H.; Zhang, R. Bioactive saponins from the fruits of *Aesculus pavia* L. *Fitoterapia*. **2011**, *82*, 1106-1109. <http://dx.doi.org/10.1016/j.fitote.2011.07.004>.

29. Lanzotti, V.; Termolino, P.; Dolci, M.; Curir, P. Pavioides A-H, eight new oleane type saponins from *Aesculus pavia* with cytotoxic activity. *Bioorganic & Medicinal Chemistry*. **2012**, *20*, 3280-3286. <http://dx.doi.org/10.1016/j.bmc.2012.03.048>.
30. Čukanović, J.; Tešević, V.; Jadranin, M.; Ljubojević, M.; Mladenović, E.; Kostić, S. Horse chestnut (*Aesculus hippocastanum* L.) seed fatty acids, flavonoids and heavy metals plasticity to different urban environments. *Biochemical Systematics and Ecology*. **2020**, *89*, 103980.
31. Zhu, C.; Peng, W.; Li, Y.; Han, X.; Yu, B. Synthesis of 3-O-(β -D-xylopyranosyl-(1 \rightarrow 2)- β -D-glucopyranosyl)-30-O-(β -D-glucopyranosyl)tamarixetin, the putative structure of aescufflavoside A from the seeds of *Aesculus chinensis*. *Carbohydrate Research*. **2006**, *341*, 1047-1051. <http://dx.doi.org/10.1016/j.carres.2006.02.036>.
32. Eastmon, R.; Gardner, R.J. [¹⁴C]Epicatechin and [¹⁴C]procyanidins from seed shells of *Aesculus hippocastanum*. *Phytochemistry*. **1974**, *13*, 1477-1478.
33. Santos-Buelga, C.; Kolodziej, H.; Treutter, D. Procyanidin trimers possessing a doubly linked structure from *Aesculus hippocastanum*. *Phytochemistry*. **1995**, *38*(2), 499-504.
34. Kimura, H.; Ogawa, S.; Sugiyama, A.; Jisaka, M.; Takeuchi, T.; Yokota, K. Anti-obesity effects of highly polymeric proanthocyanidins from seed shells of Japanese horse chestnut (*Aesculus turbinata* Blume). *Food Research International*. **2011**, *44*, 121-126. <http://dx.doi.org/10.1016/j.foodres.2010.10.052>.
35. Amiri, S.; Shakeri, A.; Sohrabi, M.R.; Khalajzadeh, S.; Ghasemi, E. Optimization of ultrasonic assisted extraction of fatty acids from *Aesculus hippocastanum* fruit by response surface methodology. *Food Chemistry*. **2019**, *271*, 762-766. <https://doi.org/10.1016/j.foodchem.2018.07.144>.
36. Castaño, J.; Rodríguez-Llamazares, S.; Contreras, K.; Carrasco, C.; Pozo, C.; Bouza, R.; Franco, C.M.L.; Giraldo, D. Horse chestnut (*Aesculus hippocastanum* L.) starch: Basic physico-chemical characteristics and use as thermoplastic material. *Carbohydrate Polymers*. **2014**, *112*, 677-685. <http://dx.doi.org/10.1016/j.carbpol.2014.06.046>.
37. Kahl, W.; Roszkowski, A.; Zurowska, A. The isolation of 6-kestose from the seeds of the horse chestnut (*Aesculus hippocastanum* L.). *Carbohydrate Research*. **1969**, *10*, 586-588.
38. Rafiq, S.I.; Singh, S.; Saxena, D.C. Effect of heat-moisture and acid treatment on physicochemical, pasting, thermal and morphological properties of Horse Chestnut (*Aesculus indica*) starch. *Food Hydrocolloids*. **2016**, *57*, 103-113. <http://dx.doi.org/10.1016/j.foodhyd.2016.01.009>.
39. Whistance, G.R.; Threlfall, D.R. Plastoquinone in leaves of *Aesculus hippocastanum*. *Phytochemistry*. **1970**, *9*, 737-738.
40. Dudek-Makuch, M.; Studzińska-Sroka, E.; Korybalska, K.; Czepulis, N.; Łuczak, J.; Rutkowski, R.; Marczak, Ł.; Długaszewska, J.; Grabowska, K.; Stobiecki, M.; Cielecka-Piontek, J.; Bylka, W.; Witowski, J. Biological activity of *Aesculus hippocastanum* flower extracts on vascular endothelial cells cultured *in vitro*. *Phytochemistry Letters*. **2019**, *30*, 367-375.
41. Salinas, F.M.; Vázquez, L.; Gentilini, M.V.; O'Donohoe, A.; Regueira, E.; Jodar, M.S.N.; Viegas, M.; Michelini, F.M.; Hermida, G.; Alché, L.E.; Bueno, C.A. *Aesculus hippocastanum* L. seed extract shows virucidal and antiviral activities against respiratory syncytial virus (RSV) and reduces lung inflammation *in vivo*. *Antiviral Research*. **2019**, *164*, 1-11. <https://doi.org/10.1016/j.antiviral.2019.01.018>.
42. Elmas, O.; Erbas, O.; Yigitturk, G. The efficacy of *Aesculus hippocastanum* seeds on diabetic nephropathy in a streptozotocin-induced diabetic rat model. *Biomedicine & Pharmacotherapy*. **2016**, *83*, 392-396. <http://dx.doi.org/10.1016/j.biopha.2016.06.055>.
43. Felipe, M.B.M.C.; de-Carvalho, F.M.; Félix-Silva, J.; Fernandes-Pedrosa, M.F.; Scortecchi, K.C.; Agnez-Lima, L.F.; Batistuzzo-de-Medeiros, S.R. Evaluation of genotoxic and antioxidant activity of an *Aesculus hippocastanum* L. (Sapindaceae) phytotherapeutic agent. *Biomedicine & Preventive Nutrition*. **2013**, *3*, 261-266. <http://dx.doi.org/10.1016/j.bionut.2012.10.014>.
44. Küçük Kurt, I.; Ince, S.; Keleş, H.; Akkol, E.K.; Avci, G.; Yeşilada, E.; Bacak, E. Beneficial effects of *Aesculus hippocastanum* L. seed extract on the body's own antioxidant defense system on subacute administration. *Journal of Ethnopharmacology*. **2010**, *129*, 18-22. <http://dx.doi.org/10.1016/j.jep.2010.02.017>.
45. Kıp, F.Ö.; Çoşkunçay, S.; Duman, F. Biosynthesis of silver nanoparticles using leaf extract of *Aesculus hippocastanum* (horse chestnut): Evaluation of their antibacterial, antioxidant and drug release system activities. *Materials Science & Engineering C*. **2020**, *107*, 110207. <https://doi.org/10.1016/j.msec.2019.110207>
46. Jarzębski, M.; Smulek, W.; Siejak, P.; Kobus-Cisowska, J.; Pieczyrak, D.; Baranowska, H.M.; Jakubowicz, J.; Sopata, M.; Białopiotrowicz, T.; Kaczorek, E. *Aesculus hippocastanum* L. extract as a potential emulsion stabilizer. *Food Hydrocolloids*. **2019**, *97*, 105237. <https://doi.org/10.1016/j.foodhyd.2019.105237>.
47. Kim, J.W.; Ha, T.-K.-Q.; Cho, H.; Kim, E.; Shim, S.H.; Yang, J.-L.; Oh, W.K. Antiviral escin derivatives from the seeds of *Aesculus turbinata* Blume (Japanese horse chestnut). *Bioorganic & Medicinal Chemistry Letters*. **2017**, *27*, 3019-3025. <http://dx.doi.org/10.1016/j.bmcl.2017.05.022>.